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ROLE OF THE SNF2 HOMOLOG, IRC20, IN YEAST GENOME MAINTENANCE

by

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Abstract

In eukaryotes, DNA is wrapped around histone proteins forming a highly compact structure, the chromatin. All DNA-based processes must occur within the complex organization of the chromatin, and this requires modulation of its structure when needed. This is accomplished by covalent histone modifications that alter histone-DNA contacts, as well as through the actions of ATP-dependent chromatin remodelers. These multi-subunit complexes play major roles in transcription regulation, replication and repairing DNA damage. This thesis aims to characterize a poorly studied member of the SWI/SNF family of ATPases/helicases, *Irc20*, from *Saccharomyces cerevisiae*. Previously, *Irc20* has been shown to be involved in recombinational repair and to possess ubiquitin ligase (E3) activity. The human homolog of *Irc20*, SHPRH, has also been implicated in repair via the poly-ubiquitylation of PCNA, the sliding clamp of the DNA polymerase. Loss of heterozygosity in the region containing the SHPRH gene is seen in a wide variety of cancers. In this study, using purified *Irc20*, we showed that it possesses DNA and nucleosome binding activities, as well as an ATP-hydrolyzing activity. However, despite homology to *Snf2* catalytic domain, *Irc20* did not have the ability to alter chromatin structure. Using point mutations in different *Irc20* domains, we identified that the increased recombination centers observed in *irc20* null mutants is dependent on both its ATPase and ubiquitin ligase activities. Consistent with this, we observed higher recruitment or retention of the recombination repair factor Rad52 at a single induced double strand break in $\Delta irc20$ mutant, suggesting a regulatory role for *Irc20* in DNA repair. Furthermore, we observed a previously unidentified function for *Irc20* in regulating the levels of the endogenous yeast 2- μ m plasmid. In *irc20* null mutant, we observed a three to four-fold increase in 2- μ m levels, forming high molecular weight forms in a manner dependent on homologous recombination. We suggest this is, at least partially, through regulating the levels of Flp1 recombinase since we observed higher levels of Flp1 in $\Delta irc20$ mutant after shutting off expression from a repressible promoter. Collectively, our results show a regulatory role for *Irc20* in recombination underlying its role stabilizing the genome and regulating the 2- μ m plasmid levels.

Keywords: *Saccharomyces cerevisiae*, *Irc20*, ATPase enzyme, ubiquitin ligase, DNA repair, 2- μ m plasmid, recombination, ubiquitin, SUMO.